





We provide the best technology & to assist you and the healthcare professional.

Building Forensics Ltd

Example Building Investigation Report

Note:

- Sections 1 to 6 are the basic report and sampling locations
- Section 7 is advised with the report to assess your mould risks
- Sections 8-9-10-11-12-13 are sometimes advised depending on health concerns and your budget
- Section 14: Conclusions
- Section 15 Recommendations

Please note

These sections are optional and may only be recommended if you have specific health concerns or your healthcare professional recommends them

We provide free guidance notes on all aspects of sampling and analysis, and will discuss your specific needs if you would like to book a call.

Executive Summary

The inspection has revealed high levels of hidden mould and airborne mould spores (genus). The symptoms you have alerted us to are likely the result of an inflammatory response from a toxigenic species, which are confirmed to be present. The analysis in section 9 confirms the presence of Mycotoxin triggers, which are also confirmed in your urine tests. The building is wet and requires professional drying. The mycotoxins identified in your medical tests are now confirmed to be present in your home.

We can provide solutions and remediation if requested.

Survey Author Jeff Charlton MCIEH CIEC- CR-WLS-CMH-Hon Fellow BDMA

Report on Building-Related Health Issues

Section 1 - Preamble

This report is based on the information you provided, an inspection of your property, and analysis and measurements taken.

No inspection can be 100% reliable, but intrusive inspection and multiple testing protocols can obtain greater accuracy. The extent and accuracy of any survey are always limited to your budget and reflect the type and number of samples taken and analysed. Any recommendations or conclusions we make must be substantiated before action.

Building-related illnesses can be challenging to treat if contamination persists. Health improvements are unlikely if exposure to inflammatory triggers exceeds the treatment benefits.

This survey may be a crucial first step in treatment. Its conclusions should be shared with your healthcare professional, and recommendations should be followed immediately.

Where biological growth has occurred, whether bacteria or mould, alive or dead, and indeed fragments, it remains a health hazard. Chemical mycotoxins and/or allergens may cause an inflammatory response decades after the initial incident. Dried fragments may be 40 times more hazardous (ref WHO) than viable growth, so even areas dried from historic water damage events may be a health risk and impossible to assess without further intrusive investigation or risk assessment.

Section 2 – Limitations of the Report

1

It should be recognised that any report can be criticised for not doing enough or for noncompliance with recognised standards. Building Forensics balances cost and required results and endeavours to provide practical and economic investigation results.

Building Forensics use recognised standards in their investigations but invariably compares target areas against unaffected regions. This can include moisture levels in air and substrates, and/or chemicals and biological activity.

Report Conclusions may indicate the presence of elevated or abnormal contamination. Generally, there is no standard for a regular or healthy home or property, and any conclusions or recommendations are based on a comparison of unaffected areas or the Building Forensics opinion.

It is, therefore, essential to recognise basic risk and hazard protocols where or if hazards and exposure routes may exist. While some moulds are considered toxigenic,

risks from allergens can create harmful synergistic effects greater than their components.

Assessments are based on probability and usually recognise the most harmful substance as the leading agent. (by CoSHH)

The bottom line is that occupant health and personal evidence of building-related illness trumps all scientific risk and hazard assessments, and our role is to support the client or patient with measurable evidence or considered sources.

Section 3 - Building Forensics Building Survey (Survey 1)

The basic survey aims to assess current and historical water damage or contamination issues, focusing on identifying possible evidence of causation. The investigation evaluated possible building construction and design defects, alterations to the building envelope and lifestyle and ventilation issues. The report culminates in conclusions and recommendations.

The report's basis is the formation of an investigation and testing of a hypothesis, which will be tested using the information provided, visual inspection, measurement, and laboratory analysis. The report emphasises building-related illnesses and the causes or likelihood of possible contamination and moisture issues. This report is not intended to be a building survey, which a RICS surveyor would generally undertake. An RICS surveyor will not typically conduct a mould or environmental survey.

This survey and inspection assess and measure areas of concern. Hidden, camouflaged, or dry areas, typically in cavities or redecorated, may not be found; however, biological sampling may identify areas of concern. Of course, this is limited to the constraints of the basic report and sampling frequency. Intrusive investigation into cavities may be required, but it is not a part of this basic investigation.

1. Report basis and considerations

1.1. Scope or Investigation Parameters

1.1.1. To assess the potential for building-related illness

1.1.2. Informed Facts –

Occupants may be suffering from a building-related illness.

The client's healthcare professional provided the following urine mycotoxin report, which indicated specific health concerns.

From our introductory survey, we identified areas to test for mycotoxins in the urine analysis.

We also identified the toxigenic species of mould likely to produce them and, most importantly, the causation. In some cases, decontamination and risk reduction are essential for treatment to be successful. **See section 14**



MycoTox Profile

OCHRATOXIN A (OTA)

Ochratoxin A (OTA) is a nephrotoxic, immunotoxic, and carcinogenic mycotoxin produced by moulds in the Aspergillus and Penicillium families. Exposure is primarily through inhalation in water-damaged buildings.

STERIGMATOCYSTIN (STG)

Sterigmatocystin (STG) is a mycotoxin closely related to aflatoxin. It is produced from several species of mould, such as Aspergillus, Penicillium, and Bipolaris. STG is considered carcinogenic, particularly in the cells of the GI tract and liver. STG has been found in the dust from damp carpets.

MYCOPHENOLIC ACID*

The Penicillium fungus produces Mycophenolic Acid (MPA), an immunosuppressant that inhibits the proliferation of B and T lymphocytes. MPA exposure can increase the risk of opportunistic infections such as Clostridia and Candida. When a woman is exposed to MPA during pregnancy, she is also at risk of miscarriage and congenital malformations.



Mould growth on the internal filter of the dehumidifier



Visible presumed mould and water staining

1.2. Building Type

- 1.2.1. Solid walls and tile
- 1.2.2. Concrete Floor

1.3. Visible and olfactory issues

- 1.3.1. No Damp Proof Course
- 1.3.2. No Air bricks
- 1.3.3. No Roof Vents
- 1.3.4. No Soffit vents
- 1.3.5. No ventilation
- 1.3.6. No trickle vents
- 1.3.7. Dead wood (mould) stored in the lounge
- 1.3.8. Decaying carpet
- 1.3.9. Visible mould on carpet
- 1.3.10. Visible mould specks on walls
- 1.3.11. Decaying window frames
- 1.3.12. A gas heater produces large quantities of moisture



Decaying carpet



No soffit vents and issues with loft ventilation

2. Dust monitoring

- 2.1.1. The airborne dust concentration is measured in g/m³ or particles defined in ppm, with sizes ranging from 0.1 microns to 10 microns. Typical equipment used is the 6-channel laser particle counter.
- 2.1.2. Fragments of mould are often in the 2.5-micron range, although spores are invariably more than 10 microns.
- 2.1.3. Particle counts are taken to assist in the development of the sampling hypothesis. The higher particle counts in association with size provide us with one indicator of possible contamination sources, and this may be where samples are taken.
- 2.1.4. The results are shown in Table 1 below

Table 1

| AREA | Particle | .3 | .5 | 1.0 | 2.5 | 5.0 | 10 |
|------------------|----------|---------------|--------------|-------|------|-----|-----|
| | Size µ | | | | | | |
| Ambient | | 16255 | 6793 | 1329 | 217 | 25 | 13 |
| Main bedroom | | 116106 | 33871 | 4135 | 636 | 99 | 53 |
| Shower | | 191649 | 54628 | 7869 | 1121 | 183 | 75 |
| Single bedroom | | 209502 | 62960 | 9012 | 1153 | 161 | 85 |
| Kitchen | | 244872 | 77360 | 11163 | 1444 | 184 | 92 |
| Hallway | | 217447 | 65223 | 9033 | 1203 | 172 | 73 |
| Lounge Piano end | | 294233 | 103434 | 13935 | 1668 | 256 | 106 |
| Lounge Table | | 297677 | 102064 | 14516 | 1612 | 247 | 93 |

2.2. Conclusions of particles

- 2.2.1. Elevated particle counts may sometimes be assumed to be sources of contamination
- 2.2.2. The highlighted readings above are elevated compared to ambient air and comparison areas
- 2.2.3. The colour-coded table is the author's subjective opinion and has no scientific value.
- 2.2.4. The air is highly contaminated, and although the content of the particulates is unknown, these are incredibly high and in my opinion a significant risk

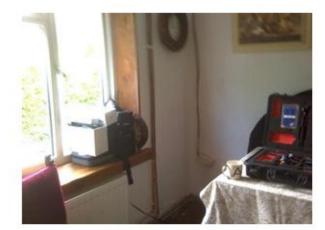
2.3. Photographic log

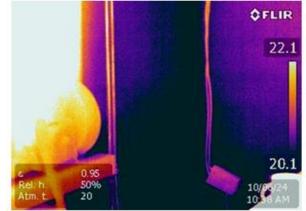


Particle counts

3. Thermal imaging survey

- 3.1. A thermal imaging camera was used to scan the building envelope and substrates to assess temperature differentials known as ΔT . These scans may identify thermal bridging and poor insulation, which can lead to dew point condensation, indicating that further investigation is required to assess possible leaks, penetrating damp, or wet materials or insulation.
- 3.2. This survey often forms the basis of the moisture mapping, but darker doesn't necessarily mean the substrate is wet; it can be cold. Darker areas in the photos can indicate cooler areas, and this may be associated with differing or missing insulation levels, dampness, air leaks, etc
- 4. The ceiling and adjacent external walls may have condensation issues







Ceiling and wall area in lounge affected by active leak from wet room leak



5. Moisture mapping

5.1.1. The moisture content of various targeted substrates is measured using moisture meters calibrated for the specific material, either with impedance (non-penetrative) or conductive pin meters.

- 5.1.2. The objective here is to assess moisture issues which may be responsible for current biological amplification. Although areas may appear dry, previous moisture may have caused hidden biological growth, which can remain allergenic or irritating until removed.
- 5.1.3. Inflammagens in cavities can be expected to leak out into the occupied spaces.
- 5.1.4. Measurement is assessed against recognised standards or the equilibrium of unaffected areas.
- 5.1.5. Equilibrium, for this purpose, is the expected homogeneous level of moisture in the same material. Where concerns are present, penetrative measurement may be required.
- 5.1.6. Materials DO NOT have to be wet or saturated for mould growth, and a boundary layer of a few molecules of moisture can exist on top of materials, which is conducive to mould growth. Mould does not grow in wet conditions and prefers damp.
- 5.1.7. Mould growth on window glass and refrigerator linings is a typical example. Although the material is non-porous, cold, and internally dry, growth still occurs on the surface. In this case, a biofilm is often the cause.
- 5.1.8. Although some materials may be dry, we also look for historically wet areas, and it must be recognised that any water damage will result in mould or bacterial amplification within 48 hours.
- 5.1.9. Biological growth (including mould) prefers damp, dark, warm areas away from UV light and air movement. Ideal growth conditions are found in the ceiling, wall and floor cavities. We may make risk assessments in the absence of complex data.

5.2. Standards of dry

5.2.1. The tables below show typical limits regarding the moisture content of various materials. Taken from British Standards PAS 64, also follows BS8201 and ASTM F2710. Further information is available in the appendix

| Structural material | MC | WME% | ERH |
|------------------------|-----|------|-----|
| Wood | 16 | 16 | N/A |
| Drywall (plasterboard) | 3.0 | 12 | N/A |
| Plaster | 0.3 | 15 | N/A |
| Brick | 1.5 | 15 | 75 |
| Concrete | 3.5 | 15 | 75 |
| Sand cement screed | 6.0 | 15 | 75 |



The floor's moisture content is 5.5, with 4.5 being an action point. This concrete floor is confirmed to be wet and may be a significant source of elevated humidity throughout the house.



A nondestructive impedance measurement of the plasterboard behind the tiles showed that water had penetrated the grouting to wet the plasterboard, resulting in hidden mould growth in the wall cavity between the shower and bedroom.



Wet plasterboard wall at 22% wme , with 12% being a trigger point for action



Infrared scanning identified possible moisture at room edges. The conductive moisture probe confirmed the floor is wet at 36% and well above the accepted 20% trigger for mould



Comparison levels indicating lower moisture issues



Comparison levels indicating lower moisture issues with the likelihood of mould in the cavity



Concrete floors wet Calibrated concrete meters are a good indicator of moisture problems, and readings should be below 4 *This may attract an additional charge*

| Area | Type of test | Reading | Concern |
|---------|-----------------|---------|---------|
| Lounge | Calcium Carbide | 5.5 | Wet * |
| Kitchen | Sleeve | 80% | Wet |

Note

These are standards, but interpretation may be required, particularly where historic water damage is present. A significant risk is where water damage in high cellulose materials has been allowed to dry naturally.

Measuring concrete, floor slabs, and screeds requires a specific and detailed investigation according to international standards.

The measurement of concrete and screed must follow a recognised protocol, which may require consideration of certain environmental factors and monitoring over 48 hours. Building Forensics will adopt an investigative approach to a lesser degree for simplicity and to contain costs. Where certified evidence is required, a separate instruction will be required.

6. Humidity ratio, also known as Specific Humidity

6.1. This is a function of relative humidity and temperature and calculates the quantity (weight) of moisture carried in the air based on g/kg of dry air. Variations between rooms and ambient conditions can indicate local moisture issues. A Thermo hygrometer with the probe is used to calculate the humidity ratio. Uncontrolled evaporation will result in moisture adsorbing into porous hydrophilic materials, possibly resulting in biological growth. Even nonporous materials can be affected by high humidity ratios, especially from the dew point condensation. The following readings with decimal points are taken directly from the meter.

| Area | Temp C | RH | Humidity Ratio |
|--------------|--------|------|-------------------|
| Ambient | 17.4 | 49.7 | 6.1 |
| Main bedroom | 18.8 | 67.7 | 9.1 |
| Shower | 18.7 | 69.8 | 9.4 |
| Single bed | 18.2 | 68.3 | 8.9 |
| Kitchen | 19.6 | 64.7 | 9.2 |
| Lounge | 17.6 | 68.5 | 8.6 |

All reasonable levels

6.2. Conclusions

- **6.2.1.** The table below shows that the property has a slightly higher specific humidity compared to ambient air, but acceptable levels
- **6.2.2.** Elevated specific humidity may be caused by lifestyle and poor ventilation. Typically, a family of four may produce 15 litres of moisture in the air per day from breathing, cooking, showers, etc.
- **6.2.3.** Drying wet clothes inside, not using cooking or bathroom exhausts, can add to this moisture loading

6.3. Dew Point Condensation

- 6.3.1. Dew point is the temperature at which warm air holding moisture condenses on colder surfaces, leaving droplets and can result in mould growth. Dew point is measured by taking the temperature of surfaces, usually external walls
- 6.3.2. External walls measured show temperatures 3 -4 degrees above dew point, condensation risk

6.3.3. Photo Log



Wall temperatures are generally 15.5, and the dew point is around 12 °c

6.4. Conclusion Low dewpoint risk



End the basic survey and start additional sampling and analysis.

Please note that we will advise and guide you to the most appropriate sampling and analysis to suit your needs and budget.

Note

While the following Total Spore Count analysis in section 7 is in addition to the preceding basic survey, we always recommend this analysis as a guide to your exposure to mould.

7. Total Spore Counts Risk Assessment

- 7.1. This survey has been developed from our experience of building-related illness and the confirmation of risks identified in the basic study 1
- 7.2. In this sampling protocol, we collect airborne spores in purpose-made, sealed cassettes for laboratory analysis by qualified mycologists. The results are compared to other areas and outside (ambient) air as a control. This type of sampling identifies the mould genus but not the species. However, the levels are an indicator of risk areas.

| Sample Number | Area collected |
|---------------|----------------|
| 1 | Bed |
| 2 | Table |
| 3 | Ambient |
| 4 | Piano |

7.3. Note

7.3.1. Mycologists cannot distinguish between Penicillium and Aspergillus. Therefore, reports are stated as Penicillium/Aspergillus, and the report is deemed a risk assessment regarding counts and types of mould. It is essential to understand that all forms of sampling and analysis have benefits and shortfalls. While this provides a risk assessment of spore counts, it cannot distinguish species or hyphal fragments.

7.4. Lab result factors

- 7.4.1. The following tables are total spore counts (viable dormant and non-viable)
- 7.4.2. The individual samples should be compared to other areas and the outside (ambient sample)
- 7.4.3. The results may be influenced by debris loading and other factors, and these findings are an integral part of the methodology. Debris loading is dust, including skin, dust mite faeces, dander and general fluff/dirt. The level of dust can obliterate the visual detection of spores when viewed under a microscope.
- 7.4.4. Debris loading is generally rated between 1 and 5, with 5 being dirty air.
- 7.4.5. A simplistic explanation is to consider a plate of peanuts with 1-2 -3 -4 -5 bags of flour dropped on top and see if you can count the peanuts through the flour.
- 7.4.6. The reality is, the dirtier the air, the more the mycologist relies on periphery counts, and this always leads to underestimation

7.5. The lab report identifies the percentage of count and the total spore count

The standard report displays a column indicating the percentage of sample fields that have been read. This means that the number of spores counted for each spore type is represented by a certain percentage of fields on which they were observed, not the percentage of the total sample.

7.6. Reporting Limits

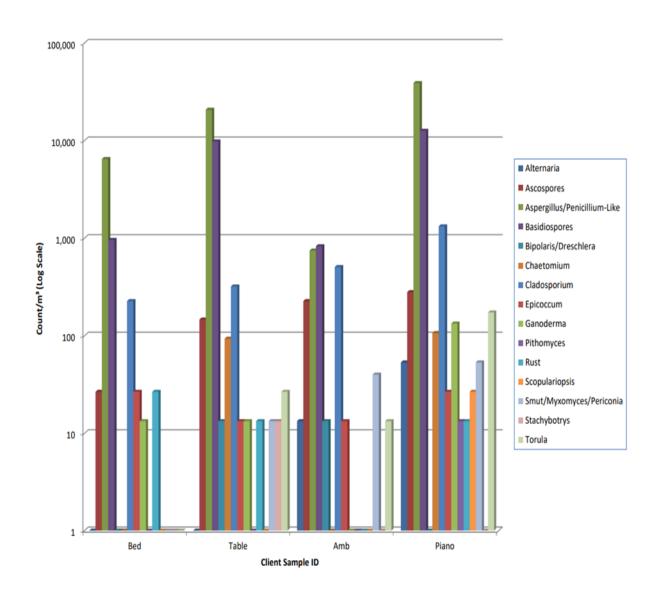
The Reporting Limit for a spore type uses the formula listed in the section above and assumes that the lowest raw count that can be detected is one.

7.7. Lab analysis

- 7.7.1. The ambient outside conditions should be compared to inside genus and spore counts (levels), including the percentage identified in the analysis. Our reports provide risk factors from the data collected.
- 7.7.2. Note the high debris loading both inside and outside the property, which may occlude visible identification; therefore, spore counts can be assumed to be higher (see note above, Table 42)
- 7.7.3. Comparison between different locations should also be considered.
- 7.7.4. The spore counts are extremely high
- 7.7.5. The lounge is grossly contaminated, but the bedroom is also bad
- 7.7.6. These identified genera may be considered toxigenic due to levels and environment

7.8. MOST IMPORTANT INFORMATION

The AMB or ambient (outside) count of Aspergillus and Penicillium is essential, as it shows 747 spores per cubic meter of air against the inside property, showing thousands of the same genus. Do not be concerned about the data gathered; we will interpret it.



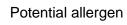
| Sample ID: | 504 | 4964-01 | | 50 | 4964-02 | | 504 | 4964-03 | | 50 | 4964-04 | |
|------------------------------|------------|----------------------|----|------------|----------------------|----|------------|----------------------|----|------------|----------------------|----|
| Client Sample ID: | | Bed | | | Table | | | Amb | | | Piano | |
| Volume Sampled (L): | | 75 | | | 75 | | | 75 | | | 75 | |
| Media: | | -O-Cell | | | -O-Cell | | | -O-Cell | | | r-O-Cell | |
| Percent of Trace Analyzed: | 100% at 60 | 0X Magnification | |
| Spore Types | Raw Count | Count/m ³ | % |
| Alternaria | _ | _ | _ | - | _ | - | 1 | 13 | 1 | 4 | 53 | <1 |
| Arthrinium | - | - | - | - | _ | - | - | - | - | - | - | - |
| Ascospores | 2 | 27 | <1 | 11 | 147 | <1 | 17 | 227 | 9 | 21 | 280 | 1 |
| Aspergillus/Penicillium-Like | 485 | 6,467 | 83 | 1,553 # | 20,707 | 66 | 56 | 747 | 31 | 2,914 # | 38,853 | 72 |
| Basidiospores | 72 | 960 | 12 | 735 # | 9,800 | 31 | 62 | 827 | 34 | 945 # | 12,600 | 23 |
| Bipolaris/Dreschlera | - | - | - | 1 | 13 | <1 | 1 | 13 | 1 | - | - | - |
| Botrytis | _ | _ | - | _ | _ | - | _ | - | - | _ | - | - |
| Chaetomium | - | _ | - | 7 | 93 | <1 | - | - | - | 8 | 107 | <1 |
| Cladosporium | 17 | 227 | 3 | 24 | 320 | 1 | 38 | 507 | 21 | 99 | 1,320 | 2 |
| Curvularia | - | - | _ | - | _ | _ | - | - | _ | - | - | _ |
| Epicoccum | 2 | 27 | <1 | 1 | 13 | <1 | 1 | 13 | 1 | 2 | 27 | <1 |
| Fusarium | _ | _ | - | _ | _ | - | _ | - | - | - | - | - |
| Ganoderma | 1 | 13 | <1 | 1 | 13 | <1 | - | - | - | 10 | 133 | <1 |
| Memnoniella | - | - | - | - | - | - | - | - | - | - | - | - |
| Nigrospora | - | _ | _ | - | _ | - | - | - | _ | - | - | - |
| Oidium/Peronospora | - | - | - | - | - | - | - | _ | - | - | - | - |
| Pithomyces | - | _ | - | - | _ | - | - | _ | - | 1 | 13 | <1 |
| Rust | 2 | 27 | <1 | 1 | 13 | <1 | - | - | - | 1 | 13 | <1 |
| Scopulariopsis | - | - | - | _ | - | - | - | - | - | 2 | 27 | <1 |
| Smut/Myxomyces/Periconia | - | _ | - | 1 | 13 | <1 | 3 | 40 | 2 | 4 | 53 | <1 |
| Stachybotrys | - | - | - | 1 | 13 | <1 | - | - | - | - | - | - |
| Torula | Ι | - | _ | 2 | 27 | <1 | 1 | 13 | 1 | 13 | 173 | <1 |
| Ulocladium | - | - | - | - | - | - | - | - | - | - | - | - |
| Unidentified Spores | I | - | - | - | - | - | I | - | - | - | - | - |
| Total Spores | 581 | 7,747 | | 2,338 | 31,173 | | 180 | 2,400 | | 4,024 | 53,653 | |
| Hyphal Fragments | 6 | 80 | | 31 | 413 | | 4 | 53 | | 31 | 413 | |
| Pollen | 5 | 67 | | 22 | 293 | | 37 | 493 | | 24 | 320 | |
| Debris Rating | | 3 | | | 3 | | | 3 | | | 3 | |
| Detection Limit | | 13 | | | 13 | | | 13 | | | 13 | |

Estimation performed due to high count.

Fungal Glossary



Typically found growing outdoors





Considered water damage indicator

Potential to produce mycotoxins

| Alternaria | | | |
|---|----------------|--|--|
| Description | Characteristic | | |
| These are a common plant pathogen involved in the decomposition of plants. In the indoor environment they are found growing on a variety of substrates including sheetrock and other building materials. They are common allergens causing hay fever or hypersensitivity reactions. | ALL ALL | | |

| Anthrinium | |
|---|----------------|
| Description | Characteristic |
| These are a plant pathogen found in soil and decomposing plant material. Not typically found growing indoors. One species has | |
| been determined to be an allergen. | |

| Ascospores | |
|---|----------------|
| Description | Characteristic |
| These are a very large group of spores that are found everywhere in nature. They are commonly found outdoors and associated with rain and moisture. Some species grow well indoors on damp materials. Ascospores have allergenic potential, however, it is species dependent. | |

| Aspergillus/Penicillium – Like | | |
|---|----------------|--|
| Description | Characteristic | |
| These are two of the most common genera in the world. They can be found everywhere in nature, both indoors and outdoors. Indoors they can be found on water damaged wallpaper, carpet, and other organic materials. They can also grow well in conditions of high humidity. Many species are allergens and a common cause of respiratory irritation. Some species are human and animal pathogens and can cause infection. | | |

| Basidiospores | | |
|--|----------------|--|
| Description | Characteristic | |
| These are primarily comprised of mushrooms and shelf fungi. They are typically found outdoors. Occasionally they are found indoors growing on any organic matter causing dry rot. Some species can be an allergen to sensitive individuals. | 2 | |

| Bipolaris/Dreschlera | | |
|---|----------------|--|
| Description | Characteristic | |
| These are a plant pathogen typically found outdoors on grasses, grains, and decaying food. Indoors they can be found on plants and building materials. They are an allergen that can affect the nose, skin, eyes and upper respiratory track. | * | |

| Botrylis | | |
|--|----------------|--|
| Description | Characteristic | |
| These are a plant pathogen typically found growing on vegetation particularly in temperate and subtropical climates. Indoors they can be found growing on plants. They are a potential allergen causing hay fever and asthma effects. | | |

| Chaetomium | |
|---|----------------|
| Description | Characteristic |
| These are typically found indoors on water damaged cellulose containing materials such as paper, sheetrock, and wallpaper. Not well studied but possible allergen with hay fever and asthma effects. | 🤧 F 🎯 |

| Cladosporium | | |
|--|----------------|--|
| Description | Characteristic | |
| One of the most common genera in both the indoor and outdoor environments. Indoors they grow well in damp environments and areas where condensation builds. They are often found on textiles, window sills, in bathrooms, and A/C systems. They are a common allergen when airborne. | | |

| Curvularia | |
|--|----------------|
| Description | Characteristic |
| Primarily found outdoors on plants and soil especially in subtropical and tropical environments. Indoors they grow on a variety of building materials. They are a common allergen causing hay fever, asthma, and allergic fungal sinusitis. | |

| Epicoccum | |
|--|----------------|
| Description | Characteristic |
| Outdoors they are found in the soil, air, and rotting vegetation. Indoors they grow well on a variety of building materials such as paper and textiles. They are a potential allergen with hay fever, asthma, and skin allergy effects. | |

| Fusarium | | |
|-------------|--|----------------|
| Description | | Characteristic |

Indoors they are typically found under very wet conditions. Some places they can be found are dust in carpet and mattresses, damp walls, wallpaper, and duct liner. They are a potential allergen causing hay fever and asthma effects.



| Ganoderma | |
|---|----------------|
| Description | Characteristic |
| These are shelf mushrooms that are typically found growing outdoors on wood causing white rot, root rot, and stem rot. They are a possible allergen at high concentration | 😝 🏆 |

| Memnoniella | |
|--|----------------|
| Description | Characteristic |
| These are mycotoxin producing spores related to and often found in conjunction with Stachybotrys. These grow well on water damaged cellulose containing building materials such as sheetrock, paper, wallpaper, and textiles. | |

| Nigrospora | |
|--|----------------|
| Description | Characteristic |
| These are typically found on decaying plant material and soil and are usually not found growing indoors. They are a potential allergen causing hay fever and asthma effects. | S |

| Oidium/Peronospora | |
|---|----------------|
| Description | Characteristic |
| These are plant pathogens that are common obligate parasites on leaves, stems, flowers, and fruits of higher living plants. | * |

| Pithomyces | |
|--|---|
| Description | Characteristic |
| These are typically found on dead leaves and stems of plants. | 1 |
| Rarely found growing indoors; however, they grow well on paper | 1997 - Carlo Ca |
| indoors given the right conditions. | |

| Rust | |
|--|----------------|
| Description | Characteristic |
| These are parasitic plant pathogens that grow on plants, grass, and trees. They are rarely found growing indoors since they require a living host, and therefore typically not found on cellulose containing building materials. They are a potential allergen causing hay fever and asthma effects. | E |
| Smut/Myxomyces/Periconia | |
| Description | Characteristic |

This is a grouping of several genera organized together in a general category that are mostly associated with living and decaying plants, wood, soil, grass, cereal crops, weeds, and flowering plants. These are rarely found growing indoors. They are a potential allergen causing hay fever and asthma effects.



| Strachybotrys | |
|--|----------------|
| Description | Characteristic |
| These are typically found indoors growing on water damaged cellulose containing building materials such as sheetrock, paper, and ceiling tiles. They are often referred to as "toxic black mold." They have the ability to produce mycotoxins which may cause a burning sensation in the mouth, throat, and nasal passages. Chronic exposure has been known to cause headaches, diarrhea, memory loss, and brain damage. | |

| Torula | |
|--|----------------|
| Description | Characteristic |
| These are typically found growing outdoors on leaves, roots, wood, and soil. Indoors they can be found growing on water damaged cellulose, paper, wicker, straw baskets and ceiling tiles. They are a potential allergen causing hay fever and asthma effects. | |

| Ulocladium | | | |
|--|----------------|--|--|
| Description | Characteristic | | |
| It requires very wet conditions and can commonly be found indoors in damp or wet areas such as bathrooms, kitchens, basements, and around windows. It grows well on cellulose- containing materials such as paper and straw, and on water- damaged building materials such as sheetrock. It is a common allergen causing hay fever and asthma symptoms. | | | |

| Unidentified Spores | | |
|---|----------------|--|
| Description | Characteristic | |
| This is a grouping of spores that cannot be categorised for various reasons. They may be weathered, disfigured, or otherwise lacking the morphological structures necessary to identify the genus. | | |

| Hyphal Fragments | | |
|--|----------------|--|
| Description | Characteristic | |
| These are branched filamentous structures with cell walls. Hyphae are somewhat analogous to stems or roots in plants, whereas the spores would be analogous to the seeds. Large quantities present may indicate an active fungal colony or active fungal growth in the structure | | |

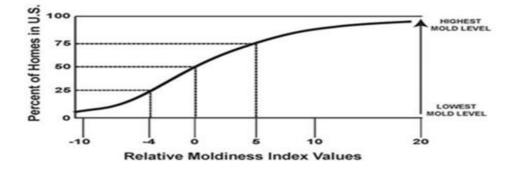
| Pollen | |
|--|----------------|
| Description | Characteristic |
| These are a fine, powdery substance produced by the anthers of seed-bearing plants, trees, grasses, flowers, and weeds. They are an allergen that causes hay fever symptoms. | |

Note

Sections 8-9 may be required where health issues are a concern. These provide additional sampling and analysis to identify the species, thereby determining the hazard levels. These are the primary indicators of building-related illness and can be used to identify mycotoxin exposure.

8. Section 8 (Hazard Assessment) is known as ERMI Settled dust and QPCR-DNA dust analysis for mould speciation (

- 8.1. The ERMI score should not be considered a risk or hazard assessment, as we usually collect dust from different areas. The risk score is derived from the presence of Group 1 moulds, which are likely to produce mycotoxins and are known as potentially toxigenic
- **8.2.** These moulds are usually present at detection levels in ambient air and are considered markers of water damage and potential health hazards.
- 8.3. While some may interpret the levels of ERMI score as a health risk, the reality is we have shown adjacent areas can have differing levels and scores (see our paper on mould sampling failures) in free advice <u>Professional Letter</u> (wpdesign.website)
- 8.4. The ERMI score has no significant value, but we interpret the actual species as a risk factor. The information regarding group 1 mould species may be of significant benefit to your medical practitioner
- **8.5.** The "Environmental Relative Mold Index" table below (ERMI) shows the typical results from 1096 homes analysed by the Environmental Protection Agency.



- 8.6. The EPA developers of ERMI categorically state its use is for professional research use only, and no significance as to risk or hazard can be gained from scores
- 8.7. No health assessments can be concluded from the ERMI score. Still, the lab results may provide important information regarding restoration and medical treatment, and the overall risk and hazards faced in the sampled areas.
- 8.8. An ERMI score of 0 would represent the average level of mould contamination (50%) of homes investigated. It should be pointed out here that a minus score

does not mean there is no health hazard or low risk. We have assessed the species, particularly in the group one section.

- 8.9. We do not advocate using the score as a risk assessment. Building Forensics assesses species from group 1 only, as a hazard assessment
- **8.10.** We have included the risk table below for those who may require information on the ERMI score.
- 8.11. After sampling over 1000 homes in the UK, we can provide a baseline of high or low, although this is not a hazard or risk assessment

| Level | ERMI Values | Interpretation | Comment |
|------------|---|----------------------------------|--|
| Q 1 | Less than - 4 | Low Relative Moldiness Index | Further investigation is not needed to determine the sources of the mold. |
| Q 2 | -4 to < 0 | Low - Medium Relative | Further investigation may be needed to determine the sources of the mold if occupants have been reactive, sensitized, |
| Q 3 | 0 to < 5 | Medium- High Relative | genetically predisposed or otherwise immuno-compromised. |
| 0.4 | 5 to < 20 | High Relative Moldiness Index | Source and cause of mold should be determined and |
| u 4 | 24 very High Relative > 20 Very High Relative Q2. | | |

Lab analysis Results - Lounge and bedroom

- 8.11.1. There are several very toxic mould species at levels over 1000fold higher than normal
- 8.11.2. The species identified can reduce your immune system, and the species in Group 1 are potentially toxigenic
- 8.11.3. The species in group 1 are most likely to be a concern to healthcare professionals who may link your symptoms to exposure.
- 8.11.4. We do not recommend using the ERMI score to assess health risks. However, the species loading is an extremely important factor in hazard assessment
- 8.11.5. Your score of 31 is in Q4 and well above the tigger of 15

| Species | Level | SE/m | g |
|----------------------------------|-------|--------|-------|
| Aspergillus flavus/oryzae | | 235 | * * |
| Aspergillus fumigatus | | 61 | * |
| Aspergillus niger | | 218 | * |
| Aspergillus ochraceus | | 4 | |
| Aspergillus penicillioides | | 12,537 | * |
| Aspergillus restrictus | | 9,311 | * * * |
| Aspergillus sclerotiorum | | 8 | |
| Aspergillus sydowii | | N.D | |
| Aspergillus unguis | | N.D | |
| Aspergillus versicolor | | 7,071 | * * |
| Aureobasidium pullulans | | 5,264 | * |
| Chaetomium globosum | | 51 | * |
| Cladosporium sphaerospermum | | 199 | |
| Eurotium (Asp.) amstelodami | | 12,436 | * * |
| Paecilomyces variotii | | N.D | |
| Penicillium brevicompactum | | 4,319 | * * |
| Penicillium corylophilum | | 139 | * |
| Penicillium crustosum | | 355 | * |
| Penicillium purpurogenum | | 17 | * |
| Penicillium Spinulosum | | 52 | * |
| Penicillium variabile | | 22 | |
| Scopulariopsis brevicaulis/fusca | | 19 | |
| Scopulariopsis chartarum | | 224 | * |
| Stachybotrys chartarum | | 5 | |
| Trichoderma viride | | 2,937 | * * |
| Wallemia sebi | | 11,003 | * |

Note

Although not as toxic as Group 1, the species identified in Group 2 are still potentially allergenic and have some toxicity.

| | | | 191 6 23 | * |
|------------------------|---|----------------------------------|----------------|---------------------------------------|
| | | | 23 | * |
| | | | | * |
| | | | | |
| | | | 8,751 | |
| | | | 317 | * |
| | | | 1,983 | * |
| | | | 1,693 | * |
| | | | 96 | |
| | | | 7,711 | * * |
| | | | 72 | * |
| n of logs G2 | 2 | | 25. | 1 |
| Ermi Results = (G1-G2) | | 31. | 1 | |
| | | n of logs G2 esults = (G1-G2) | | 96 7,711 72 n of logs G2 25. |

Environmental Relative Moldiness 31.1 Index (ERMI)

Significant results 8.12.

Building Forensics has analysed over 750 ERMI scores in the UK

Your score is almost 2.5 times higher than the UK national average •

• This HERTSMI score is 4 times the level considered safe for CIRS patients to be exposed to. (See note)

9. HERTSMI 2

9.1. This is a risk assessment of hazards identified in the QPCR-DNA sample analysis. This calculation is based on thousands of patients with varying exposures and their responses to medications, as recorded by their practitioners. The higher the HERTSMI 2 score is, the less likely the CIRS patient is to respond to treatment while those contaminants remain at high levels

HERTSMI 2 Score - Lounge and bedroom

| HERTSMI-2 Species | | Spore E. | Spore E./mg | | Weighting | |
|-----------------------|--------|-------------|-------------|----|-----------|--|
| Aspergillus penicilli | oides | 12,537 | | 10 | * | |
| Aspergillus versicol | or | 7,071 | | 10 | * * | |
| Chaetomium globosum | | 51 | | 6 | * | |
| Stachybotrys charta | arum | 5 | | 4 | | |
| Wallemia sebi | | 11,003 | | 10 | * | |
| Sample Size | 5.0 mg | HERTSMI-2 S | core = | 40 |) | |

9.2. HERTMI 2 Table of risk

| Color-coded interpretation ¹⁰ | | |
|--|---|--|
| If 10 or below | In only 1.7% of cases, re-occupancy of building following mold remediation has led to relapse of CIRS-WDB symptoms | |
| If between 11 to 15 Borderline. Further remediation and re-assessment is indicated | | |
| If greater than 15 | Re-occupancy is ill-advised until further remediation and re- assessment are conclusive. | |

9.3. Genetically close --related species may be detected in the indicator assay

| As reported | Includes |
|----------------------------------|--|
| Eurotium (Asp.) amstelodami | E. chevalieri, E. herbariorum, E. rubrum and E. repens. |
| Penicillium spinulosum | P. glabrum, P. lividum, P. pupurescens, and P. thomii. |
| Trichoderma viride | T. koningii and T. atroviride. |
| Aspergillus restrictus | A. caesillus and A. conicus. |
| Mucor amphibiorum | M. circinelloides, M. hiemalis, M. indicus, M. mucedo, M. racemosus, M. ramosissimus. |
| Rhizopus zygosporus | R. homothalicus, R. microsporus, R. oligosporus, R. oryzae. |
| Penicillium crustosum | P. camembertii, P. commune, P. echinulatum, P. solitum. |
| Aspergillus niger | Know called Aspergillus basiliensis |
| Scopulariopsis brevicaulis/fusca | Has been renamed as species of Microascus ¹⁰ |
| Wallemia sebi | W. mellicola, W. canadensis ¹¹ |

Section 10 Endotoxins

This is an addition but not usually recommended unless the budget allows, particularly where long-term mould treatments have failed, black water events/flooding have occurred, or stomach issues persist.

10. Endotoxins - Lounge and bedroom

This score is low, and endotoxins are not an issue.

| Reference Number | Locations | Result EU/mg | Q Level | | | |
|--------------------------------------|------------------|-----------------|------------|--|--|--|
| 410797-2 | Lounge + Bedroom | 24 | Q 1 | | | |
| Color-coded int pretation | | | | | | |
| If 100 or below Becommended for CIRS | | | | | | |

| If 100 or below | Recommended for CIRS. | | |
|---------------------|--------------------------|--|--|
| If 200 or below | Recommended for No CIRS. | | |
| If greater than 200 | Remediation is needed. | | |

11 Actinomycetes

Actinomycosis is believed to play a significant part in building-related illnesses and may be responsible for many symptoms, which are sometimes misdiagnosed as mould illness and CIRS. Generally, bacteria grow before mould; some studies suggest bacteria may be a higher risk factor in mould illness than mycotoxins.

11 - Lounge and bedroom

| Actino Score | 23 | |
|------------------------------|----|---|
| Pathogen Score (Q Leve) | Q4 | > |
| Black Water Score (1, Level) | Q1 | |
| Black Water Score (Level) | QI | |

| | Activo Score interpretation (Water Damage) |
|------------------|--|
| 20 or below | Indicative of a Healthy Building |
| Between 21 to 23 | Further investigation needed |
| Greater than 24 | Suggestive of Building Related Illness. |

| | Total Species | Pathogen Species | Be/mg Total | Q Level |
|----------|---------------|------------------|-------------|---------|
| Bacteria | 3,074 | 214 | 8,583,513 | Q 4 |
| Actino | 708 | 61 | | |

Summary of Bacteria's Order

| Orders Detected | Abundance B.E/mg | Families | Abundance | Fold 🔺 | Diversity | Fold 🔺 | Pathogen |
|---------------------|---------------------|----------|-----------|--------|-----------|--------|----------|
| Actinomycetales | 1,106,585 | 41 | 23 % | 0.9 | 15.7 % | 0.9 | 61 |
| Bacillales | 857,964 | 14 | 18 % | 1.4 | 5.4 % | 0.8 | 24 |
| Clostridiales | 373,495 | 25 | 8 % | 2.5 | 9.6 % | 1.2 | 36 |
| Rhizobiales | 190,651 | 13 | 4 % | 1.5 | 5.0 % | 1.1 | 1 |
| Rhodospirillales | 172,874 | 3 | 4 % | 0.8 | 1.1 % | 0.7 | 0 |
| Acidimicrobiales | 122,831 | 3 | 3 % | 5.8 | 1.1 % | 0.9 | 0 |
| Rhodobacterales | 116,230 | 1 | 2 % | 0.9 | 0.4 % | 0.6 | 0 |
| Sphingomonadales | 115,543 | 2 | 2 % | 1.8 | 0.8 % | 0.8 | 0 |
| Gaiellales | 112,873 | 1 | 2 % | 1.7 | 0.4 % | 0.3 | 0 |
| Solirubrobacterales | 94,334 | 3 | 2 % | 6.0 | 1.1 % | 1.0 | 0 |

▲ = Fold over normal top orders

Table only list 10

| Q1 Qua | artile Q2 Quartile | Q3 Quart | ile Q4 Quartile |
|---------------|---|-------------|---------------------------------|
| B.E B.E/mg | = Bacteria Equivalents = B.E/miligrams of sample | Logs ND | = Logarithms = None Detected |
| | 100 fold higher than normal. 1,000 fold higher than normal. | Ρ | = Human Pathogen |
| Distributio | alues are based on bacteria dist on of bacteria species are als are highlighted with a color code | so ranked o | n Quadriles, only elevated |

| | Actino Species Detected | | | | | |
|----|-------------------------------|--------|----------|---------|--|--|
| | Genus & Species | B.E/mg | Comments | Q Level | | |
| 1 | Actinomyces funkei | 343 | Р | | | |
| 2 | Actinomyces massiliensis | 343 | | | | |
| 3 | Actinomyces naeslundii | 343 | Р | | | |
| 4 | Actinomyces nasicola | 86 | | | | |
| 5 | Actinomyces odontolyticus | 1,803 | Р | | | |
| 6 | Actinomyces viscosus | 172 | Р | | | |
| 19 | Corynebacterium amycolatum | 5,751 | Р | | | |
| 20 | Corynebacterium appendicis | 28,669 | * | | | |
| 21 | Corynebacterium aurimucosum | 4,893 | | | | |
| 22 | Corynebacterium capitovis | 515 | | | | |
| 23 | Corynebacterium glycinophilum | 343 | | | | |
| 24 | Corynebacterium imitans | 5,064 | Р | | | |
| 25 | Corynebacterium jeddahense | 601 | | • | | |

| 26 | Corynebacterium jeikeium | 23,004 | Ρ | * | |
|----|------------------------------------|---------|---|---|--|
| 27 | Corynebacterium kroppenstedtii | 4,034 | P | | |
| 28 | Corynebacterium lactis | 429 | | | |
| 29 | Corynebacterium macginleyi | 429 | P | | |
| 30 | Corynebacterium pilbarense | 21,373 | | | |
| 31 | Corynebacterium pilosum | 86 | | | |
| 32 | Corynebacterium simulans | 6,094 | Ρ | | |
| 33 | Corynebacterium singulare | 43,347 | | | |
| 34 | Corynebacterium sputi | 172 | | | |
| 35 | Corynebacterium suicordis | 601 | Ρ | | |
| 36 | Corynebacterium tapiri | 343 | | | |
| 37 | Corynebacterium thomssenii | 7,124 | P | | |
| 38 | Corynebacterium tuberculostearicum | 255,617 | P | | |
| 39 | Corynebacterium ureicelerivorans | 4,635 | P | | |
| 40 | Corynebacterium uterequi | 172 | | | |
| 43 | Cryobacterium arcticum | 343 | | | |
| 61 | Mycobacterium aichiense | 1,373 | Ρ | | |
| 62 | Mycobacterium arabiense | 343 | | | |
| 63 | Mycobacterium cookii | 172 | | | |
| 64 | Mycobacterium duvalii | 1,373 | | | |
| 65 | Mycobacterium hippocampi | 343 | | | |
| 66 | Mycobacterium hodleri | 1,545 | | | |
| 67 | Mycobacterium holsaticum | 172 | | | |
| 68 | Mycobacterium insubricum | 86 | P | | |
| 69 | Mycobacterium iranicum | 343 | | | |
| 70 | Mycobacterium madagascariense | 3,348 | | | |
| 71 | Mycobacterium moriokaense | 1,202 | | | |
| 72 | Mycobacterium parafortuitum | 258 | | | |
| 73 | Mycobacterium rhodesiae | 258 | | | |
| 74 | Mycobacterium sediminis | 1,631 | | | |
| 75 | Mycobacterium sphagni | 343 | | | |
| 82 | Propionibacterium acidipropionici | 687 | | | |
| 83 | Propionibacterium acnes | 28,411 | P | | |
| 84 | Propionibacterium cyclohexanicum | 86 | | | |
| 85 | Propionibacterium jensenii | 172 | | | |
| 87 | Streptomyces aidingensis | 172 | | | |
| | | | | | |

| 88 | Streptomyces carpaticus | 343 | |
|----|---------------------------|-----|--|
| 89 | Streptomyces graminilatus | 429 | |
| 90 | Streptomyces nanshensis | 172 | |
| 91 | Streptomyces panacagri | 172 | |
| | | | |

| | Other Elevated Species Detected | | | | |
|---|---------------------------------|---------|----------|----------|--|
| | Genus & Species | B.E/mg | Comments | Q Level | |
| 1 | Acetivibrio cellulolyticus | 1,888 | | | |
| 2 | Acetivibrio ethanolgignens | 429 | Р | — | |
| 3 | Agromyces ramosus | 4,463 | * | | |
| 4 | Alkaliphilus oremlandii | 773 | | | |
| 5 | Amaricoccus macauensis | 12,103 | * | | |
| 6 | Ammoniphilus oxalaticus | 10,472 | * | | |
| | Anaerobacterium chartisolvens | 7,983 | | | |
| | Anaerococcus nagyae | 2,146 | | | |
|) | Anaerococcus provenciensis | 2,747 | | | |
| 0 | Anaerococcus senegalensis | 601 | | | |
| 1 | Anaerococcus vaginalis | 773 | P | | |
| 2 | Arthrobacter russicus | 2,318 | * | | |
| 3 | Bacillus asahii | 7,210 | * | | |
| 4 | Bacillus benzoevorans | 19,914 | * | | |
| 5 | Bacillus circulans | 48,411 | * | | |
| 6 | Bacillus coagulans | 5,837 | * | | |
| 7 | Bacillus foraminis | 4,120 | * | | |
| 8 | Bacillus nealsonii | 107,466 | * * | | |
| 9 | Bacillus niacini | 23,261 | * | | |
| 0 | Bacillus oceanisediminis | 27,982 | * | | |
| 1 | Bacillus psychrosaccharolyticus | 28,154 | * | | |
| 2 | Beijerinckia mobilis | 8,412 | * | | |
| 3 | Blautia faecis | 2,747 | | | |
| 4 | Blautia luti | 6,953 | | | |
| 5 | Catabacter hongkongensis | 429 | | | |
| 6 | Coprococcus eutactus | 7,124 | * | | |
| 7 | Corynebacterium accolens | 14,077 | Р * | | |
| 8 | Corynebacterium massiliense | 5,408 | * | | |

| 29 | Defluviicoccus vanus | 77,080 | * | |
|----|-----------------------------------|---------|-----|--|
| 30 | Diplorickettsia massiliensis | 11,159 | * | |
| 31 | Dorea longicatena | 2,747 | | |
| 32 | Dyadobacter hamtensis | 4,721 | * | |
| 33 | Eubacterium coprostanoligenes | 2,146 | | |
| 34 | Finegoldia magna | 11,244 | P * | |
| 35 | Friedmanniella antarctica | 4,721 | * | |
| 36 | Fusicatenibacter saccharivorans | 3,691 | | |
| 37 | Gaiella occulta | 112,873 | * | |
| 38 | Garciella nitratireducens | 1,288 | | |
| 39 | Gemmiger formicilis | 4,549 | | |
| 40 | Gracilibacter thermotolerans | 4,463 | | |
| 41 | Hydrogenispora ethanolica | 3,605 | | |
| 42 | Hyphomicrobium vulgare | 11,588 | * | |
| 43 | Intestinimonas butyriciproducens | 773 | | |
| 44 | Kocuria palustris | 83,088 | * | |
| 45 | Luteolibacter luojiensis | 7,725 | * | |
| 46 | Microlunatus ginsengisoli | 6,695 | * | |
| 47 | Microlunatus phosphovorus | 12,446 | * | |
| 48 | Nocardioides islandensis | 17,854 | * | |
| 49 | Oceanibacterium hippocampi | 22,489 | * | |
| 50 | Paenibacillus contaminans | 6,094 | * | |
| 51 | Paenisporosarcina macmurdoensis | 3,691 | * | |
| 52 | Pantoea agglomerans | 687 | Р | |
| 53 | Pantoea vagans | 2,489 | | |
| 54 | Papillibacter cinnamivorans | 515 | | |
| 55 | Peptoniphilus grossensis | 3,863 | | |
| 56 | Prevotella copri | 14,420 | * | |
| 57 | Pseudonocardia yuanmonensis | 28,326 | * | |
| 58 | Rickettsia typhi | 14,077 | Р * | |
| 59 | Rubellimicrobium mesophilum | 17,510 | * | |
| 60 | Rubellimicrobium roseum | 12,189 | * | |
| 61 | Ruminococcus callidus | 4,721 | * | |
| 62 | Siccibacter turicensis | 258 | | |
| 63 | Solirubrobacter ginsenosidimutans | 40,428 | * | |
| 64 | Sporacetigenium mesophilum | 3,004 | | |
| | | | | |

| 65 | Sporosarcina contaminans | 11,244 | * | |
|----|-----------------------------------|----------|---|--|
| 66 | Staphylococcus devriesei | 15,279 | * | |
| 67 | Staphylococcus haemolyticus | 10,987 P | * | |
| 68 | Staphylococcus saprophyticus | 8,669 P | * | |
| 69 | Symbiobacterium terraclitae | 687 | | |
| 70 | Thermoflavimicrobium dichotomicum | 6,094 | * | |
| 71 | Turicibacter sanguinis | 17,081 | * | |
| 72 | unidentified bacterium | 1,717 | * | |
| | | | | |

Cyanobacteria Species Detected Genus & Species B.E/mg Comments Q Level Aerosakkonema funiforme 343 1 2 Anabaena cylindrica 1,030 3 Anabaena flosUnclassifiedaquae 515 Microcystin Anatoxin-a, Microcystin 4 Anabaena sp 2,232 Aphanizomenon flosUnclassifiedaquae 2,918 Cylindrospermopsins, Saxitoxin 5 6 Brasilonema bromeliae 3,262 Brasilonema terrestre 4,206 7 8 Calochaete cimrmanii 773 Calothrix desertica 1,717 9 10 Calothrix elsteri 258 11 Chamaesiphon minutus 2,403 12 Chroococcidiopsis thermalis 1,030 13 Coleofasciculus chthonoplastes 86 Crinalium epipsammum 343 14 15 Crocosphaera watsonii 258 16 Cyanobacterium aponinum 86 17 Cyanobacterium stanieri 258 Cyanospira rippkae 18 343 19 Cylindrospermum siamensis 3,262 Cylindrospermum stagnale 6,781 20 21 Fischerella muscicola 86 22 Gloeobacter kilaueensis 343 23 Gloeothece membranacea 86 24 Halomicronema excentricum 944

| 25 | Halospirulina tapeticola | 172 | | |
|----|-------------------------------|-------|---|------------------------|
| 26 | Hassallia andreassenii | 343 | | |
| 27 | Hassallia antarctica | 2,489 | | |
| 28 | Iphinoe spelaeobios | 86 | | |
| 29 | Kastovskya adunca | 515 | | |
| 30 | Leptolyngbya foveolarum | 858 | | |
| 31 | Loriellopsis cavernicola | 172 | | |
| 32 | Lyngbya aestuarii | 86 | | |
| 33 | Microcystis aeruginosa | 343 | | Microcystin |
| 34 | Myxosarcina sp | 5,751 | | |
| 35 | Nodularia spumigena | 5,322 | * | Nodularin |
| 36 | Nostoc sp | 6,438 | | Microcystin, Nodularin |
| 37 | Nostoc sp | 5,064 | | Microcystin, Nodularin |
| 38 | Oscillatoria neglecta | 1,202 | | |
| 39 | Oxynema thaianum | 172 | | |
| 40 | Phormidium etoshii | 429 | | |
| 41 | Planktothricoides raciborskii | 2,232 | | |
| 42 | Pleurocapsa sp | 1,202 | | |
| 43 | Starria zimbabweensis | 1,030 | | |
| 44 | Synechococcus elongatus | 172 | | |
| 45 | Synechococcus sp | 258 | | Microcystin |
| 46 | Tolypothrix sp | 258 | | |
| 47 | Tychonema bourrellyi | 515 | | |
| | | | | |

ACTINO INDEX

Human Habitat (HH)

Soil Habitat (SH)

| Species | B.E/r | ng | |
|------------------------------------|---------|------|---|
| Actinomadura chibensis | ND | | |
| Actinomyces canis | ND | | Ē |
| Actinomyces europaeus | 86 | Р* | ī |
| Actinomyces meyeri | ND | | Ĭ |
| Actinomyces neuii | 86 | Р | ٦ |
| Actinomyces odontolyticus | 1,803 | Р* | Ĭ |
| Actinomyces turicensis | 2,146 | P ** | Ī |
| Corynebacterium accolens | 14,077 | P ** | |
| Corynebacterium amycolatum | 5,751 | Р* | |
| Corynebacterium argentoratense | ND | | Ē |
| Corynebacterium coyleae | 429 | Р* | |
| Corynebacterium falsenii | 172 | Р | Ī |
| Corynebacterium glucuronolyticum | ND | | ī |
| Corynebacterium hansenii | ND | | |
| Corynebacterium imitans | 5,064 | P | |
| Corynebacterium jeikeium | 23,004 | P ** | |
| Corynebacterium kroppenstedtii | 4,034 | Р | |
| Corynebacterium matruchotii | 86 | P | |
| Corynebacterium minutissimum | ND | | |
| Corynebacterium propinquum | ND | | Ī |
| Corynebacterium resistens | ND | | |
| Corynebacterium riegelii | 172 | Р* | |
| Corynebacterium simulans | 6,094 | P ** | |
| Corynebacterium striatum | 86 | Р | |
| Corynebacterium sundsvallense | ND | | |
| Corynebacterium tuberculostearicum | 255,617 | P ** | |
| Corynebacterium ureicelerivorans | 4,635 | Р* | |
| Corynebacterium xerosis | ND | | |
| Dermatophilus congolensis | 172 | Р | |
| Propionibacterium acnes | 28,411 | Р* | |
| Propionibacterium avidum | 858 | Р 🛎 | |
| Propionibacterium granulosum | 429 | Р* | |
| Rothia mucilaginosa | 858 | Р* | |

| Species | B.E/r | ng | | |
|--------------------------------|-------|----|----|--|
| Arthrobacter creatinolyticus | ND | | | |
| Arthrobacter crystallopoietes | 86 | Р | | |
| Brevibacterium mcbrellneri | 258 | Ρ | ٠ | |
| Brevibacterium paucivorans | 3,691 | Ρ | ** | |
| Clavibacter michiganensis | 1,288 | Ρ | ٠ | |
| Curtobacterium flaccumfaciens | 2,403 | Р | ٠ | |
| Gordonia terrae | 86 | Ρ | | |
| Nocardia higoensis | ND | | | |
| Rathayibacter tritici | ND | | | |
| Rhodococcus equi | ND | | | |
| Rhodococcus fascians | ND | | | |
| Saccharopolyspora rectivirgula | 1,202 | Ρ | | |
| Sanguibacter suarezii | 429 | Ρ | * | |

| B.E | = Bacteria Equivalents |
|-------|------------------------------|
| BE/mg | = BE/milligrams of sample |
| ND | = None Detected |
| P | = Human Pathogen |
| (*) | 5 fold higher than normal. |
| (**) | 50 fold higher than normal. |
| (***) | 500 fold higher than normal. |

Normal values is based on bacteria distribution on 1,000 US homes.

| Dominance Index (DI) | 1.1 |
|-----------------------|-----|
| Prevalence Index (PI) | 0.8 |

| Deminence Index (DI) | Lower than 2.0 | Likely safe for CIRS | |
|-----------------------|-----------------|--------------------------|--|
| Dominance Index (DI) | Higher than 2.0 | Likely not safe for CIRS | |
| Drouglance Index (DI) | Lower than 2.0 | Likely safe for CIRS | |
| Prevalence Index (PI) | Higher than 2.0 | Likely not safe for CIRS | |

Bacterial analysis results

The lab's colour-coded report shows your home to be in the fourth quantile, which is contamination above 75% worse than average.

The bacteria present are associated with cancer, including Liver and respiratory paralysis. Another bacterial species is also associated with cancer, and it can cause protein phosphatase inhibition. The levels identified from this sample are NOT very high, but you should know their presence if symptoms and/or treatment have been unsuccessful.

12. PCR-DNA Air Sampling Hazard Assessment

This sampling procedure measures the toxic loading of the air you breathe. Unlike the total spore counts, this method measures and speciates hyphal fragments and spores. This means you will discover how contaminated the air you may be exposed to is. These should only be used when health issues or high risks have been identified.

| Sample Description: | Front Top & Landing | Reporting Limit: | 3 Spores/Cubic Meter |
|---------------------------------|---------------------|----------------------------|--------------------------------------|
| Species Identification | | Spores/m3 of Air Inside | Relative Abundance (%) of Species |
| Acremonium strictum | | ND | 0.00 |
| Alternaria alternata | | ND | 0.00 |
| Anigr* | | ND | 0.00 |
| Aspergillus flavus/oryzae | | ND | 0.00 |
| Aspergillus fumigatus, Neos | artorya fischeri | 212 | 16.40 |
| Aspergillus ochraceus/ostiar | nus | ND | 0.00 |
| Aspergillus penicillicides | | ND | 0.00 |
| Aspergillus restrictus/caesille | us/conicus | 31 | 2.40 |
| Aspergillus scierotiorum | | ND | 0.00 |
| Aspergillus sydowii | | ND | 0.00 |
| Aspergillus unguis | | ND | 0.00 |
| Aspergillus ustus | | ND | 0.00 |
| Aspergillus versicolor | | ND | 0.00 |
| Aureobasidium pullulans | | 21 | 1.62 |
| Chaetomium globosum | | ND | 0.00 |
| Cladosporium cladosporioid | es svar. 1 | 13 | 1.01 |
| Cladosporium cladosporioid | | 293 | 22.66 |
| Cladosporium herbarum | | 3 | 0.23 |
| Cladosporium sphaerospern | num | 12 | 0.93 |
| Eamst* | | 15 | 1,16 |
| Epicoccum nigrum | | ND | 0.00 |
| Muc1* | | ND | 0.00 |
| Paecilomyces variotii | | ND | 0.00 |
| PenGrp2* | | 36 | 2.78 |
| Penicillium brevicompactum | /stoloniferum | ND | 0.00 |
| Penicillium chrysogenum | | ND | 0.00 |
| Penicillium corylophilum | | ND | 0.00 |
| Penicillium purpurogenum | | ND | 0.00 |
| Penicillium variabile | | ND | 0.00 |
| Pspin2* | | ND | 0.00 |
| Rhizopus stolonifer | | ND | 0.00 |
| Scopulariopsis brevicaulis/fu | ISCO | ND | 0.00 |
| Scopulariopsis chartarum | ···· | ND | 0.00 |
| Stachybotrys chartarum | | ND | 0.00 |
| Trichoderma viride/atroviride | /koningii | 631 | 48.80 |
| Wallemia sebi | • | 26 | 2.01 |
| | Tot | al Spores: 1,293 | |

| These | assays | detect | four | or mon | e species | |
|-------|--------|--------|------|--------|-----------|--|
| | | | | | | |

| Eamst | Eurotium (Aspergillus) amstelodami/chevalieri/herbariorum/rubrum/repens |
|---------|---|
| Anigr | Aspergillus niger/awamori/foetidus/phoenicis |
| PenGrp2 | Penicillium crustosum/camemberti/commune/echinulatum/solitum |
| Pspin2 | Penicillium glabrum/lividum/purpurescens/spinulosum/thomii |
| Muc1 | Mucor amphibiorum/circinelloides/hiemalis/indicus/mucedo/racemosus/ramosissimus and Rhizopus azygosporus/homothalicus/microsporus/oligosporus/oryzae |

13. Survey Mycotoxins

Survey 5 focuses on mycotoxin presence and exposure. Some of our clients have had urine sampled for a limited range of mycotoxins with American labs who often find elevated levels of Ochratoxin A, Fusarium, Sterigmatocysin, Zearalenone, etc. This can be a worry when levels are present or elevated, but the reality is that there are over 700 different mycotoxins that can affect health, and we sample for all these and the most significant mould species that produce them. Sampling in the wrong areas may provide false negative results, and survey 1 reduces that risk.

| Mould species | Mycotoxin identified in urine |
|----------------------------|-------------------------------|
| Aspergillus fumigatus | Gliotoxin |
| Aspergillus flavus | Aflatoxin |
| Aspergillus niger | Ochratoxin |
| Aspergillus versicolor | Sterigmatocystin |
| Aspergillus ochraceus | Mycophenolic Acid |
| Aspergillus penicillioides | Chaetoglobosins |
| Penicillium brevicompactum | Macrocyclic Trichothecenes |
| Chaetomium globosum | |
| Wallemia sebi | |
| Stachybotrys chartarum | |

<u>NOTE</u>

Mycotoxins, especially the seven identified above, are not the only cause of mould-related illness.

14. Conclusions

- **14.1.** The following conclusions are based on the information provided to us or gathered through the survey, coupled with laboratory analysis and monitoring equipment. While this report was written by a qualified expert in Indoor Environmental Health, you must recognise that this report is a basic, non-intrusive survey and represents a snapshot in time. You should confirm all findings before making life-changing decisions
- **14.2.** The air is extremely contaminated
- **14.3.** Surfaces are also contaminated
- **14.4.** The moisture content in the walls is elevated at the bottom compared to higher up the wall, and this may be indicative of rising damp
- 14.5. The roof has no ventilation, and the loft is an issue
- **14.6.** The levels of airborne contamination, in terms of spore counts, are incredibly high and must be reduced; the source and causation must be remediated

- **14.7.** The Gram-negative bacteria are at low levels and not considered an issue.
- **14.8.** The Gram-positive Actino levels are a little high, and I respectfully suggest you forward these results to your healthcare professional
- **14.9.** The property and its contents are extremely contaminated and pose a serious health risk.
- **14.10.** The health symptoms you described to me could, I believe, be directly attributable to building-related contamination
- **14.11.** If you have not yet engaged a healthcare professional or need some guidance, please come back to Building Forensics, and we will provide free self-help guidance and a list of appropriate healthcare support specialists

15. Recommendations

The property's contents should be risk assessed for decontamination or replacement. We can provide free guidance upon request and offer advice if you are pursuing a claim.

The roof and insulation issues must be fixed, with insulation and ventilation a priority.

The guarantee on the new floor should be reviewed, and we can provide free supportive information and guidance on this.

I recommend installing a new ventilation system, such as HRVS, as noted in section 16 below.

The property must be professionally decontaminated, and immediate risk reduction and knockdown of airborne contaminants must be considered if you remain in the property. This will be a temporary risk reduction until remediation is undertaken, as any reduced contamination will be replaced by reservoirs and defects identified in the survey.

We can decontaminate the property to various levels of risk reduction. However, costs and budget may dictate the outcome.

You can Google decontamination companies and find a variety of guaranteed results. You should review these carefully. As our client, Building Forensics will provide an unbiased, clear, and concise opinion on whoever or whatever you choose. Please also look at our guide on choosing a contractor.

I warn that any process costing under £2000 and completed in a day or two will unlikely provide any worthwhile result. We can provide you with sound advice regarding DIY monitoring and verification.

16. Decontamination Warning and Conflicts of Interest

If you have been diagnosed with CIRS or building-related illness and treatment has been unsuccessful or symptoms have worsened, you should discuss this report with your healthcare professional. If your doctor confirms the presence of potential contaminants in your home, you may require decontamination. You need a different result from the usually advertised "Mould Removal Services.' The contamination that triggers an inflammatory response is nonviable, dead, and, in fact, chemical allergens and toxins. Correct procedures are essential.

We provide free information to assist you in resolving any issues we identify. You should know that removal, NOT killing, mould is the only medically sound protocol healthcare professionals recognise.

CHAT GPT, a search engine, issued the following certificate.

You can read the complete verification by following the QR code, and also see our library on industry facts

Jeff Charlton is the **UK's leading expert** in mould investigation and remediation





April 2024